# Evaluation of Crestar® and modified Crestar programs for timed insemination in lactating Egyptian buffaloes (Bubalus bubalis) under intensive production system

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The current study was conducted on a total of 204 Egyptian, lactating buffalo cows. These animals were in the second or third parity, of good body condition scores and apparently healthy. The animals were raised in intensive production system on a private farm. The buffalo cows were allotted into three groups, two of these groups were experimental and the third was a control group. The first experimental group included 30 buffalo cows were undergo ovulation control by Crestar® a subcutaneous ear implant (3mg norgestamet) plus Crestar® injection i.m. (3mg norgestamet + 5mg estradiol valerate ) at zero day. At the 7<sup>th</sup> day of implantation, PGF2α was injected i.m., then Crestar® implant was removed at the 9<sup>th</sup> day with injection of PMSG 400 iu. Timed insemination was conducted 56 hrs later. The second experimental group (24 buffalo cows) was treated by the same program, moreover they injected with GnRH at the time of insemination. The third group (150) buffalo cows was bred naturally and used as a control group. For serum progesterone assay blood samples were collected from the animals of the two experimental groups at day 0, 7 and 9 of the Crestar program. The buffalo cows of the experimental groups were closely observed for estrus signs and were rectally palpated at the time of insemination for detection of the internal estrus changes. At day 50 post insemination all animals were rectally palpated for pregnancy diagnosis. The result of the current study revealed that the visibility of estrus signs were 20 %, 16.7% and 22 % for the first, second and third group respectively. Pregnancy rate was much higher in the second group associated with the injection of GnRH at the time of insemination. Two animals of the second group were carrying twins (11 %). Serum level of progesterone was significantly higher in the 7<sup>th</sup>day in comparison with those recorded for 0 and 9th day.

Buffaloes were introduced into Egypt from India, Iran and Iraq approximately during the middle of the 7th Century. It is the most important and popular livestock for milk production in Egypt. Population size is 3 717 000 and resemble 55 % of total bovine population in Egypt (El-Kirabi, 1995; FAO 2003).

Reproductive efficiency is the primary factor affecting productivity and is hampered in female buffalo by inherent late maturity, Poor estrus expression, Distinct seasonal reproductive patterns and Prolonged intercalving intervals (Madan, 1988; Madan and Raina, 1984). Moreover, prolonged postpartum acyclicity (absence of ovarian cyclic activity) and anestrum (absence of overt estrous signs) are major sources of economic

loss to buffalo breeders (Singh et al., 2000; El-Wishy 2007).

Improvement of reproductive efficiency in buffalo requires the identification of specific limiting factors under a given situation and the development and field testing of strategies for improvements and interventions that are sustainable with available local resources (Perera, 1999).

In Egypt, more than 90 % of buffaloes are raised as small holder's. The traditional farmer possesses only one to four buffaloes and usually no bull; hence, there is little opportunity for behavioral interaction among estrus animals ((El-Kirabi, 1995)). Artificial control of the estrous cycle has provided an efficient means of increasing

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the reproductive capacity of buffalo by obviating the need for frequent visual inspections (Madan, 1988).

Prostaglandins have been used to induce estrus in buffalo, but they work if a corpus luteum is present and therefore they can be useful in subestrus animals, having a synchronizing more than an inducing effect (Dhalival et al., 1988 Chohan et al., 1995; Sahasrabudhe and Pandit, 1997; Awasthi et al., 1998; Kharche and Srivastava, 2001). The use of gonadorelin (GnRH), given by multiple injections or in microencapsulated form, did not and moreover efficacious administration times are not of practical use (Shah et al., 1990; Fateh et al., 1999; Takkar et al., 1999). More useful and efficacious have been the treatments using progesterone associated with gonadotrophin or gonadorelin (Zicarelli and Boiti, 1982; Rao and Sreemannarayana, 1983; Singh et al., 1983, 1984, 1988; Borghese et al., 1993; Shanker et al., 1999; Hattab and Osman, 2000; Hattab et al., 2000).

Borghese *et al.*, (1993) ,reported that, Italian Buffalo cows raised under intensive production system when treated with subcutaneous implants of Norgestomet + PMSG and Buserelin (GnRH-analogue) released by subcutaneous osmotic pump; or Progesterone + Buserelin i.m. have been able to reduce the intercalving interval and increase the fertility of the herd out of the breeding season. Better results have been obtained using progesterone - releasing intravaginal device (PRID) associated with PMSG and prostaglandin (Zicarelli *et al.*, 1994).

The aim of the current study was to evaluate the use of Crestar® and modified Crestar program for control of ovulation and timed insemination in the lactating Egyptian buffaloes under intensive production system in well managed dairy.

### Material and methods

Animals and management. This study was conducted from December, 2004 to December 2005, using lactating (n = 204) second or third-parity, Egyptian buffaloes (*Bubalus bubalis*) from the herd maintained on El-Komy Farms (Kilo 138 Cairo-Alexandria Desert Road- Egypt). The buffaloes used were of good body condition scores, weighed from 500-600 kg and free from any apparent anatomical or reproductive disorders. All of these buffaloes were in the breeding period (60-120) days post partum). The buffaloes were

kept under loose housing conditions in clean, hygienic opened yard system with sandy-muddy flooring, asbestos roofing equal to 30 % of floor square area, and sufficient space for the free movement of the animals (40 square meters for each animal). All buffaloes were fed a ration with total mixed ration (TMR), consisting of concentrates (maize grain, Soya been cake, wheat bran), roughages (either Barseem or wheat straw), a mineral mixture, and salt .The concentrate/ roughage ratio was 55 / 45 %. Fresh water was available ad libitum. These animals were milked twice daily by milking machines with special bucket adopted for buffaloes .The average daily milk production was 10 kg /head /day. Calf rearing was depending on natural suckling of colostrum for five days and then isolated from his dam and raised on natural buffalo milk with bucket feeding. Synchronization of ovulation. Buffaloes were randomly assigned into three groups, two experimental groups (group A and B) and control one (group C). The two experimental groups were collected randomly from different yards in two yards free from bulls. The first experimental group (group A) included 30 buffalo cows and were undergo ovulation control by Crestar® (Intervet, Boxmeer, The Netherlands ). Crestar® consists of 2 components: an ampoule of 2 ml injection containing estradiol valerate (5 mg) with norgestomet (3 mg) and a silicone ear implant containing3 mg norgestomet (17a-acetoxyl-l 1 gmethyl- 19-nor-pregn-4-ene-3, 20-dione). At zero day, the injection was administered im and the implant was inserted subcutaneous (SC) at the outer edge of the ear in all animals. After 9 days the norgestomet implants were removed. Two days before implant removals 2ml Prosolvin (a synthetic analogue of PGF2α containing 15 mg of luprostinol, Intervet, Boxmeer, The Netherlands) was injected i/m. At the ninth day, 400 I.U. PMSG (Folligon, Intervet, Boxmeer, the Netherlands) was injected i.m. Artificial insemination conducted, 56 hours after implant removal.

The second experimental group (group B) of 24 buffalo cows was treated with modified Crestar program. It was including Crestar® plus the injection of  $100~\mu g$  GnRH (Gonadorelin Intervet, Boxmeer, the Netherlands) at the time of insemination. The third group (Group C), including 150 buffalo cows was bred naturally and used as a control group.

Traits  Animal groups	No.	Visible estrus signs % (N)	Internal estrus changes						
			Vulval changes % (N)	Uterine changes % (N)	Ovarian changes % (N)	Pregnancy rate % (N)	Twinning % (N)	Still birth % (N)	Abortion % (N)
Group A	30	20 % (6)	60 % (18)	73 % (22)	80 % (18)	60 % (18)	-	_	_
Group B	24	16.7 % (4)	54 % (13)	79 % (19)	83 % (20)	75 % (18)	11 % (2)	5.5 % (1)	5.5 % (1)
Group C	150	22 % (33)	_	_	_	56 % (84)	2.3 % (2)	4.7 % (4)	3.5 % (3)

Table 1: Estrus activities and reproductive traits in timed insemination and naturally bred Egyptian buffalo cows.

Group A: Timed insemination with Crestar  ${\bf @}$ 

Group B: Timed insemination with modified Crestar

**Group C: Naturally bred Animals** 

Table 2: blood serum Progesterone profile in controlled ovulation Egyptian buffalo (Mean ± SE ng / ml).

Progesterone	At 0 day N (54)	At 7th day N (54)	At 9th day N (54)
Mean ± SE	$3.75\pm0.53^{a}$	6.75± 0.73 <sup>b</sup>	2.68± 0.32 a
Min. Value	.09	2.09	0.14
Max. Value	12.07	18.07	7.01

Means within the same raw with different alphabetical were significantly different at  $p \le 0.01$ .

Artificial insemination and natural mating. Animals of the two experimental groups (group A and B) were artificially inseminated by a good quality of frozen semen produced by a world wide company imported from Italy. Meanwhile animals of the third group (control group of 150 buffalo cows) were bred naturally by good fertile bulls (one bull per 25 buffalo cows in separate yards).

**Estrus activities.** Animals of the two experimental groups were palpated per rectum to assess the internal estrus changes at the time of insemination (presence of follicle > 1 cm, tonic uterus and presence or absence of mucous vaginal discharge). Estrus detection for the animals of the control group was performed twice a day in early morning and at the afternoon by visual observation of well trained workers.

Collection of blood samples. Blood samples (5 mL for determination of progesterone concentrations) were collected by jugular venipuncture into polystyrene tubes at 0, 7 and 9 days from insertion of Crestar implant.. Blood samples were chilled on ice, transported to the laboratory and centrifuged at 3,000 x g for 15 minutes. Serum was kept at -20" C until assayed. Progesterone assay was measured using

FERTIGENIX PROG-EASIA by BIOSOURCE EUROPE S.A.Kit according to Matthiews (1986).

**Pregnancy diagnosis.** Pregnancy diagnosis of the two experimental groups was assessed at 50 days after artificial insemination and confirmed at 90 days by rectal palpation of the uterine contents. Pregnancy rates were calculated for animals that were still pregnant at 90 days.

Regarding the control group (group C), first rectal examination was performed after 40 days for animals with detected estrus and then at 90 days .Meanwhile animals without detected estrus were checked for pregnancy every three weeks. Statistical analyses. Differences between serum progesterone concentrations between the two experimental groups during treatment were analyzed using repeated measure ANOVA using PC-STAT (1985).

## **Results**

#### Responce to synchronization.

Visible estrus signs. On the day of AI, 20% of animals in group A and 16.7 % of group B showed estrus behavior. Meanwhile the percentage of animals showed estrus signs were 22% in group C at the time of natural mating (table 1). Frequent urination is the most observed symptom of estrus and bellowing. Mucus vaginal discharge in quietly

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sitting animals was observed. Many buffaloes showed clear or transparent cervical mucus at the onset of estrus.

**Internal estrus changes.** Edema of the vulva was observed in 60 and 54 % in the animals of group A and B respectively .Uterine tone and ovarian changes were more obvious in group B than group A (table 1).

Pregnancy rates and twinning. As shown in table 1, closed values of pregnancy rates were recorded for the animals treated with Crestar® program and the control group (60 and 56 % respectively). Meanwhile a higher value (75 %) of pregnancy rate was recorded for animals treated with modified Crestar program. Two animals of group B were carrying twins; abortion was recorded for one of them at the 7<sup>th</sup> month of gestation and the second buffalo gave still birth.

concentration Serum of progesterone. concentration Summaries for serum progesterone were presented in table 2.A highly significant value  $(6.75\pm 0.73 \text{ ng/ml})$  was recorded for all treated animals in group A and group B at the 7th day of insertion of Crestar implant, with mean values averaged from 2.09 and 18.07 ng / ml). Meanwhile closed values were recorded for all animals in day 0 and day 9 0f treatments (3.75± 0.53 and  $2.68 \pm 0.32$  ng / ml, respectively).

#### **Discussion**

The failure of buffaloes to show overt signs of estrus, together with the wide variation in duration of estrus, are major constraints to the proper adoption of AI for genetic improvement in buffaloes (Bruselli et al., 2001). There is a requirement, therefore, to identify an estrus synchronization treatment that results in reliable and consistent synchronization of stage of the estrous cycle and associated with a relatively high pregnancy rate when combined with fixed - time AI in buffaloes. Previous studies in estrus synchronization in dairy buffaloes undergoing commercial milking have tended to utilize small numbers of animals and it has been difficult to identify a preferred estrus synchronization protocol (Hattab et al., 2000).

In this regard, an important feature of the present study was the relatively large numbers of buffaloes used to evaluate the progesterone (Crestar®) and a new modification of this program by injection of GnRH at the time of insemination in synchronization of ovulation and timed –

insemination. It was found that treatment with Crestar® and modified Crestar program both achieved good synchrony in ovulation in buffaloes as judged by the high pregnancy rates especially with the modified Crestar program. The overall pregnancy rate after synchronization with Crestar® was 60%. This study was consistent with previous reports in Italian buffaloes (Rao and Rao, 1983; de Araujo et al., 2002). They reported pregnancy rates 0f 50% and 56%, respectively. Meanwhile the present study was inconsistent with the study of Neglia et al. (2003) who reported ,pregnancy rate 28% was synchronization with PRID® .The later study was conducted during the transition to seasonal anestrus for Italian Mediterranean buffaloes in Southern Italy. Moreover it was proven that the use of PRID together with PMSG treatment is able to induce fertile estrus in non-cycling buffalo heifers (Barile et al., 2001; Pacelli et al., 2001). The high pregnancy rate in the current study in treated and control groups may be referred to the high body condition scores and all of these animals were not suckling animals. Body condition score (BCS) plays an important role in the reproductive performance of post-partum buffalo cows. Baruselli et al., (2001). They reported that, first post-partum estrus was influenced by BCS at calving; cows with high BCS had an earlier first post-partum estrus and a shorter service period than cows with lower BCS.

Suckling significantly increases the interval from parturition to first estrus in buffalo. Jainudeen et al., (1983) found that in Malaysian Swamp buffaloes that suckled their calves, had showed a significant increase in the interval from parturition to first ovulation than milked buffalo cows. An earlier resumption of ovarian activity in milked rather than suckled buffaloes was found by El-Fouly et al., (1976). These authors report that only 38 percent of suckled buffaloes restored ovarian activity within 90 days from parturition. This result disagreed with that reported by Janudeen et al., (1983). The extension of anoestrus period due to calf suckling is also reported by Usmani et al., (1990). They found a post-partum estrus cyclicity resumption delayed by three to four weeks due to the practice of let buffaloes be suckled by their calves, before each milking, to stimulate milk let down. Arya and Madan (2001) also found a longer interval from parturition to

first observed estrus and a longer service period in suckled than weaned buffaloes

In the current study ,a high percentage of pregnancy rate was recorded for the second group (group B). This is consistent with Rastegarnia *et al.*, (2004) who concluded that injection of 100 microgram of Gonadorelin is the most effective dose to induce ovulation in river buffalo (Bubalus bubalis).

Regarding progesterone concentration, in the current study, it was  $3.75\pm0.53$ ,  $6.75\pm0.73$  and  $2.68\pm0.32$  ng / ml at zero,  $7^{th}$  and the  $9^{th}$  day respectively. This value agreed with the report of Hattab *et al.*, (2000); Campanile *et al.*, (2005) .Peak progesterone values have been recorded about 15 days after estrus (Bachlaus *et al.*, 1979; Arora and Pandey, 1982; Takkar *et al.*, 1983).

In the current study the highly significant increase was recorded after 7 days of Crestar implant, this is consistent with Ahmed *et al.*, (1977), who reported that, the first significant increase in progesterone concentration occurs about 7 days after estrus in normal cycling buffaloes.

#### Conclusion

The use of Crestar® as progesterone implant was useful as an aid for ovulation control and timed insemination in the Egyptian buffalo cows .However GnRH injection at the time of insemination significantly improved the pregnancy rate.

#### References

- Ahmed, A.; Agarwal, S. P.; Agarwal, V. K.; Rahman, S. A. and Laumas, K. R. (1977): Steroid hormones: Part II. Serum progesterone concentration in buffaloes. Indian J. Exp. Biol., 15:591–593.
- **Arora, R. C. and Pandey, R. S. (1982):** Changes in peripheral plasma concentrations of progesterone, estradiol-17 beta, and luteinizing hormone during pregnancy and around parturition in the buffalo\_Bubalus bubalis. Gen. Comp. Endocrinol., 48:403–410.
- **Arya, J. S. and Madan, M. L. (2001):** Post partum reproductive cyclicity based on ovarian steroids in suckled and weaned buffaloes. Buffalo J., 17 (3): 361-369.
- Awasthi, M. K.; Tiwari, R. P. and Pangaonkar, G. R. (1998): Induction of oestrus and fertility with low dose of prostaglandin F2 alpha in suboestrus buffaloes. Indian J. Anim. Sci., 68 (10): 1049-1050.
- Bachlaus, N. K.; Arora, R. C.; Prasad, A. and Pandey, R. S. (1979): Plasma levels of gonadal hormones in cycling buffalo heifers. Indian J. Exp. Biol., 17: 823–825.
- Barile, V. L.; Galasso, A.; Marchiori, E.; Pacelli, C.; Montemurro, N. and Borghese, A. (2001): Effect of PRID

- treatment on conception rate in Mediterranean buffalo heifers. Livest. Prod. Sci., 68: 283-287.
- Baruselli, P. S.; Barnabe, V. H.; Barnabe, R.C.; Visintin, J. A.; Molero-Filho, J. R. and Porto R. (2001): Effect of body condition score at calving on postpartum reproductive performance in buffalo. Buffalo J., 17 (1): 53-65.
- Borghese, A.; Barile, V. L.; Campanile, G.; Esposito, L. and Pacelli, C. (1993) :Induzione dell'estro in bufale acicliche. Nota I. Percentuale di ciclicità. [Oestrus induction in acyclic water buffaloes. Note I. Cyclicity rate]. In: Atti 5° Meeting Nazionale "Studio della Efficienza Riproduttiva degli Animali di Interesse Zootecnico", Bergamo, Italy, 30:125-129.
- Campanile, G.; Di Palo, R.; Neglia, G.; Vecchio, D.; Gasparrini, B.; Prandi, A.; Galiero, G. and D'Occhio, M. J. (2007):Corpus luteum function and embryonic mortality in buffaloes treated with a GnRH agonist, hCG and progesterone. Theriogenol., 67(8):1393-1398.
- Chohan, K. R.; Iqbal, J.; Choudhary, R. A. and Khan, A. H. (1995): Oestrous response and fertility in true anoestrus buffaloes following hormonal treatment during summer. Pakistan Vet. J., 15:6–8.
- de Araujo, A.; Berber, R. C.; Madureira, E. H. and P. S. Baruselli, (2002): Comparison of two Ovsynch protocols (GnRH versus LH) for fixed-timed insemination in buffalo (*Bubalus bubalis*), Theriogenol., 57:1421–1430.
- **Dhalival, G. S.; Sharma, R. D. and Singh, G. (1988):** Efficacy of prostaglandin F2 alpha administration for inducing oestrus in buffalo. Theriogenol., 29 (6): 1401-1406.
- **El-Fouly, M. A.; Kotby, E. A. and El-Soubhy, H. E. (1976):** Post partum ovarian activity in suckledand milked buffaloes. Theriogenol., 5: 69-79.
- **El-Kirabi, F. (1995):** Buffalo population and production in Egypt. Buffalo Newsletter, 3: 8-9.
- El-Wishy, A. B. (2007): The postpartum buffalo. II. Acyclicity and anestrus. Anim.Reprod.Sci., 97(3-4):216-36. FAO (2003): www. FAO.org /DAD-IS.
- Fateh, M.; Dhaliwal, G. S. and Sharma, R. K. (1999): Clinical efficacy of GnRH analogue (buserelin) and oestradiol benzoate treatments in anoestrus buffaloes. Indian J. Anim. Sci.,69 (5): 310-312.
- **Hattab, S.A. and Osman, R. A. (2000)**:Use of Norgestomet and oestradiol valerate injection with norgestomet implant and/or PMSG for induction of oestrus in anoestrous buffaloes. Vet. Med. J., Giza, 48 (1): 135-143.
- **Hattab S. A.; Kadoomz, A. K.; Palme R. and Bamberg E..(2000):** Effect of Crestar<sup>TM</sup> on estrus synchronization and the relation ship between fecal and plasma concentration of progesterone in buffalo cows. Theriogenology; 54(7): 1007-17.
- **Jainudeen, M. R.; Bongso, T. A. and Tan, H. S. (1983):** Postpartum ovarian activity and uterine involution in the suckled swamp buffalo (Bubalus bubalis). Anim. Reprod. Sci., 5: 181-190.
- **Kharche, S. D. and Srivastava, S. K.(2001)**:Fertility following treatment of suboestrus buffaloes with prostaglandin F2a. Indian J. Anim. Reprod., 22 (2): 170 171. **Madan, M. L. (1988)**: Status of reproduction in female buffalo. In: Buffalo Production and Health: a compendium of latest research information based on Indian studies. ICAR Publication, New Delhi, India, pp. 89–100.

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**Madan, M. L. and Raina, V. S.(1984):**Fertility and performance of buffaloes under tropical conditions. In: 10<sup>th</sup> Int. Congr. on Anim. Reprod. and Artificial Insemination, Illinois vol. 2, pp. 142.1–142.4.

- Matthiews, C. (1986): Serum progesterone level as an aid in the diagnosis of ectopic pregnancy. Obstetrics Gynaecol., 68:390-394.
- Neglia, G. B.; Gasparrini, R. D.; Palo, C. D.; Rosa, L. Zicarelli, N. and Campanile, G. (2003): Comparison of pregnany rates with two estrus synchronization protocols in Italian Mediterranean buffalo cows, Theriogenol., 60., 125–133
- **Perera, B. M. (1999):** Reproduction in water buffalo: comparative aspects and implications for management. J.Reprod.Fertil.Suppl., 54:157-68.
- Pacelli, C.; Barile, V. L.; Lenza, R.; Terzano, M. G.; Montemurro, N. and Borghese, A. (2001): Comparison of two different doses of PMSG on conception rate in Mediterranean buffalo heifers treated with PRID. In: Proc. Sixth World Buffalo Congress, Maracaibo, Venezuela, 160-165.
- PC-STAT (1985): Statistical programs .Coded by Mohn Rao, Khathlen Blane and Marc Zonnenberg, Univ. of Georgia.
- **Rao, A. R. and Rao, C. C. (1983):** Synchronisation of oestrus and fertility in buffaloes with a progesterone releasing intravaginal device. Vet. Rec., 113:623.
- Rao, A. V. N. and Sreemannarayana, O.(1983): Induction of ovulatory oestrus and fertility in non cycling buffaloes with norgestomet during low breeding season. Therigenol., 19
- Rastegarnia, A.; Niasari-Naslaji, A.; Hovareshti, P. and Sarhaddi, F., Safaei, M. (2004): The effect of different doses of gonadorelin on ovarian follicle dynamics in river buffalo (Bubalus bubalis). Theriogenol., 62(7):1283-91.
- Sahasrabudhe, S. A. and Pandit, R. K. (1997):PGF2a induced oestrus in suboestrus Murrah buffaloes during the summer. Indian J. Anim. Sci., 67: 513-514.
- Shah, S. N. H.; Willemse, A. H. and Van De Wiel, D. F. M.(1990): Reproductive performance of Nili-Ravi buffaloes after a single injection of GnRH early post partum. Tropic. Anim. Health and Prod., 22 (4): 239-246.
- Shanker, U.; Agarwal, S. K.; Kumar, S. and Mohan, G. (1999): Oestrus response and fertility using progestagen ear implant in noncyclic buffalo (Bubalus bubalis). Indian J.Vet.

- Res., 8 (1): 54-58.
- Singh, G.; Singh, G. B.; Sharma, R. D. and Nanda, A. S. (1983): Experimental treatment of summer anoestrus buffaloes with norgestomet and PRID. Theriogenol., 19 (3): 323-329.
- Singh, G.; Singh, G. B.; Sharma, R. D. and Nanda, A. S. (1984):Ovulation and fertility after PRID, PRID+GnRH and GnRH in anestrous buffaloes. Theriogenol., 21 (6): 859-867.
- Singh, G.; Dhaliwal, G. S.; Sharma, R. D. and Biswas, R.K. (1988): Treatment of summer anestrous buffalo (Bubalus bubalis) with progesterone releasing intravaginal device plus pregnant mare serum gonadotropin. Theriogenol., 29 (5): 1201-1206.
- Singh, J.; Nanda A. S. and Adams G. P.( 2000): The reproductive pattern and efficiency of female buffaloes, a review. Animal Reprod. Sci., 593–604.
- **Takkar, O.P.; Singh, M. and Verman, P.N.(1983):** Progesterone levels vis a vis anoestrum in buffaloes concurrent with profile during stages of oestrus cycle. Indian J. Dairy Sci., 36: 125–128.
- Takkar, O. P.; Singh, N.; Kaur, J. and Chaudhary, K.C. (1999): Augmentation of postpartum reproductive efficiency by use of GnRH in buffaloes, Indian J. Anim. Reprod., 20 (1): 9-11.
- **Usmani, R. H.; Dailey, R. A. and Inskeep, E. K.** (1990):Effects of limited suckling and varying prepartum nutrition on postpartum reproductive traits of milked buffaloes. J. Dairy Sci.,73: 1564-1570.
- Zicarelli, L. E. and Boiti, C.(1982): Risultati preliminari sull'impiego in primavera di spirali vaginali con progesterone nella bufala. [Preliminary results on the use of a progesterone intravaginal device in Italian buffalo cows during the spring season]. In: Atti II Convegno Internazionale sull'Allevamento Bufalino nel Mondo, Caserta, Italia, 29 Settembre 2 Ottobre: 412-
- Zicarelli, L.; Campanile, G.; Esposito, L.; Di Palo, R.; Boni, R.; Spadetta, M.; Montemurro, N.; Pacelli, C.; Borghese, A.; Barile, V.L.; Terzano, G.M.; Annicchiarico, G.; Allegrini, S.; De Benedetti, A.; Malfatti, A.; Lucaroni, A. and Todini, L.(1994):Anaestro e induzione dell'estro in bufale acicliche. [Anoestrus and oestrus induction in acyclic buffaloes]. Agricoltura Ricerca, 153: 25-40.

# تقييم برنامج غرسة كرستار و كرستار المعدل كإحدى وسائل توحيد التلقيح الموقوت في الجاموس المصرى الحلاب تحت ظروف الإنتاج المكثف.

أجريت هذه الدراسة على عدد ٢٠٠٤ جاموسة حلابة موزعة على موسمى الإنتاج الثانى والثالث في إحدى المزارع الخاصة في مصر وكانت هذه الحيوانات في حالة صحية وجسمانية جيدة ، هذا بالإضافة إلى خلوها الظاهر من الأمراض التناسلية ، قسمت هذه الحيوانات إلى ثلاث مجموعات كالتالى :- المجموعة (أ) و تشتمل على ٣٠ حيوان و تم معاملتها بوضع غرسة الكرستار التى تحتوى على هرمون البروجستيرون تحت سطح الجلد الخارجي للأذن و في نفس الوقت حقنت هذه الحيوانات بعن الكرستار و هو عبارة عن ٢ ملل تحتوى على خليط من هرموني البروجستيرون و الاستروجين ، في اليوم السابع تم حقن هذه الحيوانات بالبروستاجلاندين F2 و بعدها بيومين ، أي في اليوم التاسع تم إزالة الغرسة من الأذن و في نفس الوقت تم حقن هذه الحيوانات بعد ٥ ساعة من إزالة الغرسة ، المجموعة (ب) و تشتمل على ٢٠ حيوان تم معاملتها بنفس المعاملة السابقة مع حقن الهرمون المنابه للغذة المنافق في الحظائر دون أيية المجموعة (ج) و تشتمل على ٢٠ حيوان و هي عبارة عن مجموعة قياسية و لقد تركت هذه الحيوانات مع الطلائق في الحظائر دون أية معاملات هرمونية ، تم أخذ عينات دم من هذه الحيوانات في اليوم الأول و اليوم السابع واليوم التاسع من المعاملات لقياس مستوى هرمون البروجستيرون في مصل الدم، لوحظت هذه الحيوانات في اليوم الأول و اليوم السابع واليوم التاسع من المعاملات لقياس مستوى يوما من التلقيح لتحديد العشار من عدمه ، كشفت نتيجة الدراسة الحالية بأن روية إشارات الشياع الظاهر في المجموعات أ ، ب ، ج هي ٢٠ ، البروجستيرون سجلت أعلى القراءات في اليوم السابع من الغرسة مقارنة بساعة الصفرو اليوم التاسع ، نستنتج من هذه الدراسة ١ – برنامج غرسة الكرستار هو برنامج فعال لإستخدام التلقيح الموقوت في الجاموس وهو ما يساعد كثيرا في إجراء عملية التحسين الوراشي ٢ – حقن غرسة الكرستار هو برنامج فعال لإستخدام التلقيح الموقوت في الجاموس وهو ما يساعد كثيرا في إجراء عملية التحسين الوراشي ٢ – حقن المهرون المنبه للغذة النخامية أثناء عملية التلقيح يحسن من نتائج العشار .