

Antimicrobial activity of some cephalosporins with special reference to their effects on body weight and immune response to Newcastle disease vaccine in fayoumy chicks

I. A. Radwan¹, Abeer M. Radi^{2*}

¹Microbiology and ²Pharmacology Departments, Faculty of Veterinary Medicine, Beni-Suef University, Beni-Suef, 62511, Egypt.

The susceptibility of the most common bacterial pathogens, namely *E. coli*, *P. mirabilis* and *Ps. aeruginosa* which were isolated from egg incubators and yolk sacs of randomly selected one day old Fayoumy chicks to three selected cephalosporins (cephradine, ceftiofur and cefquinome) were studied. The minimum inhibitory concentration (MIC), the minimum bactericidal concentration (MBC) of the tested drugs and the effect of these antibiotics on the body weight gain, mortality and immune response against Newcastle disease (ND) vaccine of the same breed of chicks were also estimated. The tested organisms were sensitive to ceftiofur and cefquinome whereas *E.coli* and *ps. aeruginosa* were found to be resistant to cephradine. The results showed that mortalities were higher in control and cephradine treated groups, while it was lower in the ceftiofur and cefquinome treated groups. On the other hand, the lowest mean body weight was recorded in control group (155.7±6.55 gm) followed by ceftiofur treated group (162.5±2.06 gm) and the highest mean body weight was recorded in cefquinome treated group (183.5±1.66 gm, $p < 0.01$) at 30 days of age. The study revealed that the tested antibiotics not exert any immune suppressive effect against (ND) vaccine.

Colibacillosis refers to localized or systematic infection caused entirely or partially by *Escherichia coli* (*E.coli*), including colisepticaemia, coligranuloma, air sac disease, cellulites, swollen head syndrome, peritonitis, salpingitis, osteomyelitis, synovitis, omphalitis and yolk sac infection (Barnes and lozano, 1994).

Proteus mirabilis (*P. mirabilis*) was isolated from dead-in-shell chicken embryos (Orajaka and Mohan, 1985). The organism can penetrate egg shell and survive within the egg (Al-Aboudi *et al.*, 1988). On the other hand, *Proteus mirabilis* infection mainly causes mortality in young chicks up to 4 weeks of age with suppurative osteomyelitis (Venkanagouda *et al.*, 1996).

Pseudomonas organism can cause localized or systemic diseases in young and growing poultry, invade fertile eggs causing death of embryos and newly hatched chicks. *Pseudomonas aeruginosa* (*P. aeruginosa*) can be highly virulent causing 100 % mortality in experimentally inoculated chickens Lin *et al.*, (1993), and from 0 to 90% in broiler chicks (Walker *et al.*, 2002).

Cephradine is 1st generation cephalosporin

that can be administered orally and used successfully in treating infection of the respiratory tract and soft tissue. The gram-negative aerobic enteric bacilli (*E.Coli*, *P. mirabilis*, *Salmonella spp.*, and *Shigella spp.*) were reported to be highly susceptible to the first generation cephalosporins, (Quintilani *et al.*, 1982).

Ceftiofure is a broad spectrum 3rd generation cephalosporin approved for veterinary use to treat a variety of Gram-negative, Gram-positive bacterial pathogens and anaerobic pathogens including *Pasterilla spp.*, *Streptococcus spp.*, *Staphylococcus spp.*, *salmonella spp.* and *Escherichia coli* (Brown *et al.*, 1991; Salmon *et al.*, 1996; Tragesser *et al.*, 2006). Ceftiofur was recommended for the control of *P.multocida* infection in balady chickens and for control of terminal bacterial infection in one day old broiler chickens (El-Naeneey and lotfy, 2000).

Cefquinome is the 4th generation cephalosporin developed for use in veterinary medicine. It had in vitro and in vivo efficacy against a wide range of gram-negative and gram-positive bacterial pathogens, (Limbert *et al.*, 1991; Rohdich *et al.*, 2009).

The present study was carried out to.

Determine the susceptibility of *E.coli*, *P. mirabilis* and *Ps. aeruginosa* isolated from egg incubators and yolk sac of randomly selected day

* Corresponding author. Tel.: +20 082 2322066;

Fax: +20 082 2327982

E-mail address: a.radi@bsu.edu.eg

(Abeer M. Radi)

old Fayoumy chicks to three selected cephalosporins (cephradine, ceftiofur and cefquinome).

Determine the effect of these antibiotics on chicks' performance.

Studied the effect of the selected antibiotics on chicks' immune response to (ND) vaccine.

Materials and methods

Drugs.

Cephradine (velocef)[®]. It was supplied as pure powder for injection from Bristol-Myers Squibb Company, Cairo, Egypt.

Ceftiofur sodium (Excenel)[®]. It was supplied as sterile powder (1 gram vial) for injection from Pharmacia & Upjohn Company, Cairo, Egypt.

Cefquinome sulphate (Cobactan)[®]. It was obtained as suspension 2.5% for injection from Intervet International Company Cairo, Egypt.

Samples. A total of 10 samples in the form of cracked shells, dust and fluffs were collected from hatcheries, as well as eighty day old Fayoumy chicks were randomly selected for isolation of bacterial pathogens from their yolk sacs. Forty day old chicks were randomly selected for collection of blood samples for application of Hemagglutination Inhibition test for (ND) maternal antibodies.

Media for bacterial isolation and identification. Bacterial fluid and solid media were used for isolation of different bacterial pathogens, as each swab was inoculated into tetrathionate broth, tryptone soya broth and MacConkey broth. All fluid media were incubated at 37 °C for 18 hours. A loopfull from each fluid medium was seeded onto the surface of the following solid plating media; salmonella-shigella agar, Tryptone soya agar and MacConkey-bile salt-lactose agar media, followed by aerobic incubation at 37 °C for up to 72 hours. The obtained colonies were identified morphologically and smears were taken stained with Gram's staining technique and identified biochemically according to Collee *et al.*, (1996).

Experimental chicks and grouping. A total number of 1400 one day-old Fayoumy chicks were divided into 4 equal groups; each of 350 chicks. All chicks were floor reared in El-Azab project for poultry production at Fayoumy Governorate. Feeding and lighting programmes were applied according to the catalogue of the project.

In vitro antimicrobial sensitivity test. The bacterial isolates from the egg incubators and

egg yolk were examined for sensitivity against three cephalosporins (cephradine, ceftiofur and cefquinome).

Cephradine, ceftiofur and cefquinome sensitivities were determined using the disc and agar diffusion method as described by Collee *et al.*, (1996) as follows. A portion of a single bacterial colony was selected and inoculated into 4 ml Muller Hinton broth and incubated at 37 °C for 18 hours. After incubation the concentration of the M.O was equal to 1×10^8 cfu/ml. The culture was flooded onto the surface of well-dried Muller Hinton agar plates. The plates were tipped and excess of fluid was removed with a pipette after being sure that entire surface was covered with inoculum. The bacterial culture was allowed to settle for 15 minutes at 37 °C. The antimicrobial discs of cephradine and ceftiofur (30 µg/disc) and cefquinome (30 µg/well) were aseptically overlaid. The plates were allowed to stand 30 minutes at room temperature, after application of the discs. Then after overnight incubation at 37 °C, the plates were examined for inhibition zones. The interpretation of the results were carried out according to Collee *et al.*, (1996).

Minimum inhibitory and bactericidal concentrations [(MIC) and (MBC)]. According to Collee *et al.*, (1996) the drugs were dissolved in sterile distilled water to obtain concentrations of 10 mg ml⁻¹, 1 mg ml⁻¹, 0.1 mg ml⁻¹ as stock solutions.

From the 10 mg ml⁻¹ solution 256 µl, 128 µl, 64 µl, 32 µl were taken and added to tubes containing 20 ml Muller Hinton broth to obtain the following dilutions of antibiotics (128, 64, 32 and 16 µg ml⁻¹, respectively).

From the 1 mg ml⁻¹ solution 160 µl, 80 µl and 40 µl were taken and added to tubes of Muller Hinton broth to obtain the following dilutions of (8, 4, 2 µg ml⁻¹).

From the 0.1 mg ml⁻¹ tube 200 µl, 100 µl, 50 µl and 25 µl were added to Muller Hinton broth tubes to obtain dilutions of (1, 0.5, 0.25 and 0.125 µg ml⁻¹).

Each tube was inoculated with 100 µl of 1×10^8 cfu/ml (prepared by using MecFarland opacity tube) suspension of each of the microorganisms isolated from the egg incubators and chicks.

All tubes were incubated at 37 °C for 24 hours.

At the end of incubation period the tubes were examined visually for turbidity.

The tubes which has no visible growth indicate the MIC points.

To determine the MBC, a loopfull from the tubes which showing no visible growth were subcultured onto Muller Hinton agar and incubated at 37 °C for 24-48 hours. After incubation of the subculture plates they were examined for growth. The tube containing the lowest concentration of antibiotic that fails to yeiled growth on the subculture plate was regarded as containing MBC of antibiotic for the tested strain.

Vaccination programme. Chicks groups were received the following viral vaccines.

At 1 day old. Marek's virus vaccine s/c in the neck 0.2 ml/chick.

At 2 day old. Infectious bronchitis living attenuated vaccine in drinking water.

At 6 days of age. ND (Hitchner B1) living attenuated vaccine in drinking water.

At 9 days of age. Infectious bursal disease vaccine in drinking water which repeated at 19 days of age.

At 18 days of age. ND (Lasota) vaccine in drinking water which was repeated at 28 days of age.

Experiment NO 1 (The effect of subcutaneous injection of the tested drugs on the chicks performance). In this experiment the effect of subcutaneous injection of three doses (at one, 16 and 26 days of age) of cephradine, ceftiofur and cefquinome was studied on the performance (general health condition, body weight, feed intake, food conversion rate and mortality rate) of 1400 one day old chicks about 27 gm body weight. The chicks were divided into 4 groups each of 350 chicks. The 1st group was injected with cephradine at adose of 50 mg kg⁻¹ b.wt (Oishi *et al.*, 1976), the 2nd group injected ceftiofur (10 mg/kgb.wt.), Aziza *et al.*, (1998), whereas the 3rd group injected cefquinome (2 mg kg⁻¹ b.wt.) Block (1996) and the 4th group was was kept as a non treatrd control. The chicks of each group was weighed at 1, 7, 14, 21, 30 days of age.

Relative growth rate and feed conversion ratio were calculated according to Crampton and Lioyd (1959) as the following:

$$\text{Relative growth rate} = \frac{(w_2 - w_1)}{(w_1 + w_2) \div 2} \times 100$$

Where:

w1= body weight at the beginning of the period.

w2 = body weight at the end of the period.

The feed conversion rate was measured by dividing the amount of food consumed, in a

certain period, by the gain in weight at the same period, expressed in the same weight units.

Experiment NO 2 (The effect of the tested drugs on chicks immune response against ND vaccine). Serum samples were collected at the first day by scarifying 40 chicks, then at the 10th, 19th and 29th day of age by scarifying 15 chicks from each group and collection of their blood for serum. Collected sera were subjected to for heamagglutination inhibition test (HI) by the conventional microtitre method according to Calnek (1979).

Statistical analysis. The mean and standerd deviatons were calculuted as described by Snedecor (1969), the F- test was carried out by using INSTAT program.

Results and Discussion

Bacterial isolates from examined hatchery samples, 6 samples were found to harbour bacterial pathogens from 10 samples, (2 *Ecoli*, 3 *ps. aeroginosa* and 1 *p. mirabilis* isolate) and 22 bacterial pathogens were isolated from the yolk sac of day old chicks, and identified as (9 isolates *Ecoli*, 8 isolates *ps. aeroginosa*, 3 isolates *p. mirabilis* and 2 isolates were spore forming aerobic gram- positive bacilli).

These findings nearly coincide with the results obtained by Choudhury *et al.*, (1993) who isolated *E.coli* from 57 (67.04 %) of infected yolk sacs on 58 farms. Barnes and Lozano (1994) revealed that egg transmission of pathogenic *E.coli* is common and can be responsible for high chick mortality. Radwan and Hassan (2004) experimentally induced 68 % embryonic death using *p.mirabilis* through egg shell.

E. coli and *ps. aeroginosa* were found to be resistant and *p .mirabilis* was found to be sensitive to cephradine. These finding resemble those obtained by Lacey *et al.*, (1983); Gakuya *et al.*, (2001).

The tested organisms was sensitive to Ceftiofur, this is similar to the result obtained by (El -Naenaey and lotfy 2002) who recorded that in chicks 39 out of 40 tested strains of *E.coli* were sensitive to ceftiofur, for *Ps. aeruginosa* the activity percentage of 80 % was recorded to ceftiofur as 4 strains were sensitive from the tested 5 isolates, all tested isolates of *proteus* species from chicks were completely sensitive. Huang *et al.*, (2009) reported that *E.coli*, *salmonella* and *p. multocida* isolates form poultry were sensitive to ceftiofur. On the other hand our study not agrees with the result obtained by (Deshpande *et al.*, 2000) who found

that the isolates of *E.coli* which produce extended spectrum β -lactamase (ESBL) were resistant to ceftiofur. This may be due to the acquired resistance of *E.coli*. Also (Walker *et al.*, 2002) recorded that antibiotic sensitivity tests showed that *p. aeruginosa* isolates obtained from hatcheries and broiler chicks were resistant to ceftiofur.

The result of our investigations proved that *E. coli*, *P. mirabilis* and *ps. aeruginosa* were sensitive to cefquinome. This finding agreed with that reported by Sheldon *et al.*, (2004) and Rohdich *et al.*, (2009).

MIC and MBC of cephradine for the tested *E. coli*, *P. mirabilis* and *Ps. aeruginosa* were 32, 128 and $>128 \mu\text{g ml}^{-1}$ and 64, 128, $>128 \mu\text{g ml}^{-1}$, respectively (Table 1).

The MIC and MBC of ceftiofur for *E. coli*, *P. mirabilis* and *ps. aeruginosa* investigated in this study were 0.5, 16, $>128 \mu\text{g m}^{-1}$ and 0.5, 16 and $>128 \mu\text{g ml}^{-1}$ respectively. These results similar to that recorded by (Deshpande *et al.*, 2000), the MIC of ceftiofur and cefquinome for *E.coli* were (0.06-2 and $\leq (0.03 -1 \mu\text{g ml}^{-1})$), respectively but resistant strains of *E.coli* exert MIC $>32 \mu\text{g ml}^{-1}$ for both drugs. Meyer *et al.*, (2008) recorded that the MIC for *E.coli* in foals was $\leq (0.5-1 \mu\text{g/ml})$.

This study revealed that MBCs for ceftiofur is similar to its MICs against most tested strains strongly suggest that ceftiofur exerts bactericidal effect. This result confirmed the findings of Franklin (1992) and Klein *et al.*, (1996); they reported that ceftiofur sodium exerts bactericidal effect on tested isolates at concentration equal to or at most one doubling dilution above MIC.

The MIC and MBC of cefquinome for the tested bacteria *E. coli*, *P. mirabilis* and *Ps. aeruginosa* investigated in this study were 0.5, 1, 16 $\mu\text{g ml}^{-1}$ and 1, 16 and $>128 \mu\text{g ml}^{-1}$, respectively. This finding similar to that obtained by Limbert *et al.*, (1991) who recorded that the MIC for *E. Coli* ($<0.006-0.781 \mu\text{g ml}^{-1}$), *P. mirabilis* (0.024-0.39 $\mu\text{g ml}^{-1}$), *ps. aeruginosa* (0.391-50 $\mu\text{g ml}^{-1}$). Also the MIC for *E Coli* and *P. mirabilis* was less than or equal to 0.5 $\mu\text{g ml}^{-1}$, while MIC for *ps. aeruginosa* 8 $\mu\text{g ml}^{-1}$ and MBC for most species except Enterobacter were within a dilution of MIC, Chin *et al.*, (1992); Murphy *et al.*, (1994). Cefquinome had MIC value for *E.coli* $<0.06 \mu\text{g/ml}$ in cattle by Sheldon *et al.*, (2004) and Thomas *et al.*, (2006).

Performance of different groups of chicks under study. As shown in Tables (2 and 3), mortalities were higher in control (53 chicks)

and cephradine treated group (33) and lower in the other two groups (21 and 20) in ceftiofur and cefquinome treated groups, respectively). Regarding total feed intake, the control non-treated group showed the highest quantity of feed intake (632.78 gm/chick in 30 days) followed by cefquinome treated group (593.21 gm/chick). On the other hand, the lowest mean body weight was recorded in control group (155.7 gm \pm 6.55) followed by ceftiofur treated group (162.5gm \pm 2.06) and the highest mean body weight was recorded in cefquinome treated group (183.5 gm \pm 1.66), ($p < 0.01$) in 30 days. Mean weight gain was highest in cefquinome treated group (156.5) and lowest in control group (128.7). The excellent feed conversion rate was obtained in the third cefquinome treated group (3.79) followed by ceftiofur treated group (4.09) mean while it was (4.18) and (4.92) in cephradine and control groups, respectively. Relative growth rate was highest (148.69) in cefquinome treated group and lowest (140.88) in control treated group (Table 3).

Egg-born diseases are transmitted from the infected dam to newly hatched offspring by means of fertile eggs. Some disease agents are carried inside the shell as a result of shedding into the egg prior to the addition of shell membrane. Others are carried out on the shell or penetrate from the shell surface through pores after the egg is laid (Radwan and Hassan, 2004). The ability of ceftiofur and cefquinome to reduce mortality rate were detected in this study and the efficacy of both drug were also evident by improved mean body weight gain, feed intake and feed conversion. This previous data is supported by Alexander, (1985) who reported that after the lapse of the acute phase of the infection the drugs improve weight gain in consequence of an increased feed intake and increased absorption of nutrients.

The used chicks had ND maternal antibodies titers of $\log_2 6$. Serum samples were collected from all groups at 10, 19 and 29 days of age and the mean antibody titer of each group was showed in Table (4).

It was clear that ceftiofur treated group recorded non significant decrease antibody titer against ND vaccine, mean while cefquinome treated group recorded non significant increase antibody titer.

Conclusion. From the previous study we could concluded that:

Cephradine could be used for treatment of potential infections with *Proteus mirabilis*.

Table (1): Minimum inhibitory and bactericidal concentrations {(MIC) and (MBC)} of the tested drugs.

Tested M.Os	Cephadrine		Ceftiofur		Cefquinome	
	MIC ($\mu\text{g ml}^{-1}$)	MBC ($\mu\text{g ml}^{-1}$)	MIC ($\mu\text{g ml}^{-1}$)	MBC ($\mu\text{g ml}^{-1}$)	MIC ($\mu\text{g ml}^{-1}$)	MBC ($\mu\text{g ml}^{-1}$)
<i>E. coli</i> (1 isolates)	32	64	0.5	0.5	0.25	1
<i>P. mirabilis</i> (1 isolates)	128	128	16	16	1	16
<i>Ps.aeruginosa</i> (1 isolates)	>128	>128	>128	>128	16	>128

Table (2): Effect of cephradine, ceftiofur and cefquinome on mortalities and performance in chicks.

Group (350 chicks/group)	Mortality per number (350 chicks)	Feed intake (gm) per chick	Mean body weight (gm) per chick n =80 chicks	Mean weight gain (gm) per week	Feed conversion rate (week)	Relative growth rate
7 days of age						
1 st group (Cephadrine)	8 chicks	87.7	57.3±0.47	30.25	2.89	71.8
2 nd group (Ceftiofur)	9	87.98	54.75±1.49	27	3.25	65.06
3 rd group (Cefquinome)	6	86.48	58.5±1.5	31.5	2.74	73.68
4 th group (Control group)	27	91.33	57.5±0.645	30.3	3.014	71.7
14 days of age						
1 st group (Cephadrine)	13	149.5	94±2.81	36.8	4.06	48.6
2 nd group (Ceftiofur)	6	158.9	92.4±1.43	36.4	4.4	49.06
3 rd group (Cefquinome)	10	156.86	100.6±1.79	42.1	3.72	52.9
4 th group (Control group)	13	148.4	92.75±0.36	35.25	4.2	46.93
21 days of age						
1 st group (Cephadrine)	2	179.1	130.4±1.46*	36.2	4.95	32.3
2 nd group (Ceftiofur)	1	160.4	132.1±2.27**	39.7	4.04	35.3
3 rd group (Cefquinome)	2	166.1	135.2±0.71***	42.1	3.94	34.6
4 th group (Control group)	3	206.7	123.6±1.03	31.6	6.54	24.73
30 days of age						
1 st group (Cephadrine)	10	152.06	163±3.93	32.6	4.66	22.3
2 nd group (Ceftiofur)	5	146.96	162.5±2.06	30.4	4.83	20.6
3 rd group (Cefquinome)	2	183.77	183.5±1.66**	40.8	4.51	25
4 th group (Control group)	10	186.35	155.7±6.55	32.1	5.8	22.1

* P < 0.05

** P < 0.01

*** P < 0.001

Table (3): Comparison between different groups of chicks demonstrating important items of performance.

Performance	Group			
	1 st group (Cephadrine)	2 nd group (Ceftiofur)	3 rd group (Cefquinome)	4 th group (control non treated group)
Total mortalities	33	21	20	53
Total average Feed intake (gm)/chick in 30 days	568.36	554.24	593.21	632.78
Mean body weight (gm) at 30 days of age	163±3.93	162.5±2.06	183.5±1.66**	155.7±6.55
Mean body gain in 30 days	136	135.5	156.5	128.7
Food conversion rate at 30 days of age	4.18	4.09	3.79	4.92
Relative growth rate at 30 days of age	143.16	143	148.69	128.7

** P < 0.01

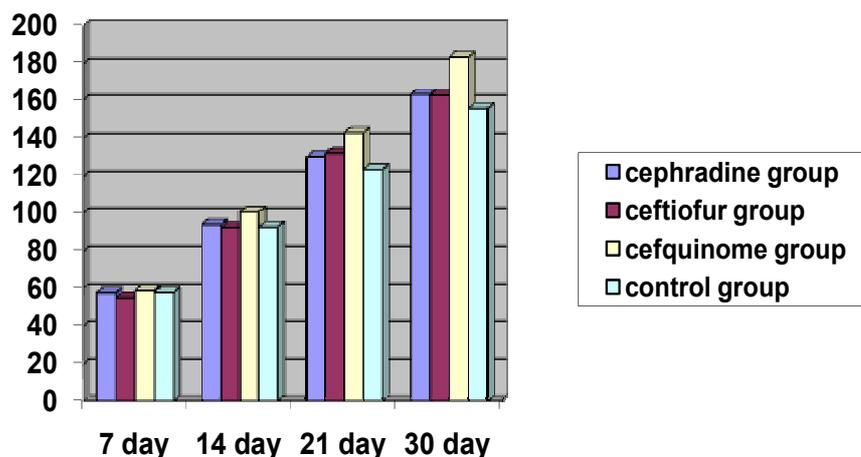


Fig. (1): Mean body weight in the different chicks groups.

Table (4): Effect of cephradine, ceftiofur and cefquinome on ND Mean HI log₂ of antibody titers.

Age per days	Group			
	Cephradine group	Ceftiofur group	Cefquinome group	Control non treated group
One day age	6	6	6	6
At 10 days of age	11	11	9.85±0.40	10±0.45
At 19 days of age	9.57±0.68	6.714±0.61	9±0.44	8.16±0.6
At 29 days of age	8.33±0.45	5.7 ±0.39	9.1±0.655	7.33±0.65

Non significant variation ($p > 0.05$).

Cefquinome and ceftiofur could be used for treatment of potential infections with *Esherchia coli*, *Proteus mirabilis* and *pseudomonas aeruginosa* microorganisms.

Mortalities were lower in chicks treated by ceftiofur and cefquinome while the highest mean body weight was recorded in cefquinome treated chicks.

Cephradine, ceftiofur and cefquinome did not exert any immunosuppressive effect against Newcastle disease vaccine thus they could be used safely before vaccination without adverse effect on the immune response of chickens.

References

Al-Aboudi, A. R.; Shinawa, I. M.; Hassen, A. A. and Al-Sanjary, R. B. (1988): Penetration rate of proteus organisms through egg shell membranes at different temperature. Iraqi J. Vet. Sci., 1: 1-8.

Alexander, F. (1985): An introduction of vet. pharm. 4th Ed. Longman (London and NewYork).

Aziza, M. M. Amer; Eisha, M. M. Fahim and Rawia, K. Ibrahim (1998): Effect of aflatoxicosis on the kinetic behaviour of ceftiofur in chickens. Res. Vet. Sci., 65(2): 115-118.

Barnes, H. J. and Lozano, F. (1994): Colibacillosis in poultry. In Pfizer veterinary practicum, Pfizer Animal Health. Leeyes summit MO. 45 pp.

Block, C. Von (1996): Pharmacokinetics of the cephalosporin antibiotic cefquinome in sows at different stages of the reproductive cycle. Inaugural-Dissertation, Tierarztliche Fakultat, Ludwig-Maximilian Universitat, Munchen, Germany.

Brown, S. A.; Yancey, R. J. and Stef, J. (1991): Ceftiofur sodium: Antimicrobial activity of parent ceftiofur and metabolites. Acta Veterinaria Scandinavica; 87:95-97.

Calnek, B.W. (1979): Haemagglutination-inhibition antibodies against an adenovirus (virus127) in white pekin ducks in the United States. Avian Dis., 22(4):798-801.

Chin, N.; Gu, J.; Fang, W. and Neu, H. (1992): Invitro activity of cefquinome, a new cephalosporin, compare with other cephalosporin antibiotics. Diagn. Microbial Infect. Dis., 15(4):331-337.

Choudhury, B.; Chanda, A.; Dasgupta, P.; Dutta, R.K.; Lila-saha; Sanatan-Bhuin; saha, L. and Bhuin, S.(1993): Studies on yolk sac infection in poultry, antibiogram of isolates and correlation between in-vitro and in vivo drug action. Ind. J. Anim. Hlth., 32(1):21-23.

Collee, C. J.; Barrie, P. M.; Andrew, G. F. and Anthony S. (1996): Cited in Mackie & McCartney Practical medical microbiology. 4th ed. 151-178. Newyork EclinBurgh London.

Crampton, E.W. and Lioyd, L.E. (1959): Fundamentals of nutrition. W.H. Freeman and company. Sanfranciso and London.

Deshpande, L.; Pfaller, M. A. and Jones, R.N. (2000): In vitro activity of ceftiofure tested against clinical isolates of *Escherichia coli* and *Klebsiella pneumoniae* including

- extended spectrum-beta lactamase producing strains. Int. J. Antimicrob. Agents, 15(4): 271-275.
- El-Naeneey, E.Y. and Lotfy, O. O. (2002):** Experimental and field trials to evaluate the antibacterial action of ceftiofur sodium (Excenel) in chickens. Vet. Med. J., Giza. 48(2): 207-216.
- Franklin, A. (1992):** The in-vitro bactericidal activity of Danofloxacin and ceftiofur against respiratory pathogens in cattle. Proceedings 17th world Buiatrics Congress, 3:214-217.
- Gakuya, F. M; Kyule, M. N; Gathura, P. B. and Kariuki, S. (2001):** Antimicrobial resistance of bacterial organisms isolated from rats. East. Afr. Med. J., 78(12):646-649.
- Huang, T. M.; Lin, T. L. and Wu, C.C. (2009):** Antimicrobial susceptibility and resistance of *Escherichia coli*, *Salmonella* spp. and *Pasteurella multocida* isolates. Avian Dis., 53(1):89-93.
- Klein, L. k; Yancey, R. J.; Case, C. A. and Salmon, S. A. (1996):** Minimum inhibitory concentration of selected antimicrobial agent against bacteria isolated from 1-14 day old broiler chicks. J. Vet. Diag. Invest., 8 (4):494-495.
- Lacey, R. W.; Gillian, L. H.; Vickil, I.; Luxton, D. E. A. and Trotter, I. S. (1983):** Double-blind study to compare the selection of antibiotic resistance by Amoxycillin or Cephradine in the commensal flora. The lancet, 322(8349): 529-532.
- Limbirt, M.; Isert, D.; Klesel, N.; Markus, A.; Seeger, K.; Seibert, G. and Schrin, E. (1991):** Antibacterial activities in vitro and in vivo and pharmacokinetic of cefquinome (HR111V), a new broad-spectrum cephalosporin. Antimicrob. Agents Chemoth., 35(1):1419.
- Lin, M.Y.; Cheng, M. C.; Huang, K. J. and Tsai, W. C. (1993):** Classification, pathogenicity, and drug susceptibility of hemolytic Gram-negative bacteria isolated from sick or dead chickens. Avian Dis., 37:6-9.
- Murphy, S. P.; Erwin, M. E. and Jones, R. N. (1994):** Cefquinome (HR11V) in vitro evaluation of a broad-spectrum cephalosporin indicated for infections in animals. Diagn. Microbial. Infect. Dis., 20:49-55.
- Oishi, M.; Nishizuka, K.; Motoyama, M.; Ogawa, T. and Tanaka, M. (1976):** Ophthalmic application of intravenous cephradine. Jpn. J. Antibiot., 29(11):958-962.
- Orajaka, L. J. E. and Mohan, k. (1985):** Aerobic bacterial flora from dead-in shell chicken embryo from Nigeria. Avian Dis., 29:583-589.
- Quintilani, R.; French, M. and Nightingale, G. M. (1982):** First and second generation cephalosporins. Antimicrobial Therapy. Med. Clin. North Am.; 66: 183-197.
- Radwan, I. A. and Hassan, W. H. (2004):** Comparative exclusion of proteus mirabilis causing cloacal prolapse and embryonic chicken deaths. Beni-Suef Vet. Med. J. XIV. (1):29-37.
- Rohdich, N.; Zschiesche, E.; Heckerroth, A.; Wilhelm, C.; Leendertse, I. and Thomas, E. (2009):** Treatment of septicaemia and severe bacteria infections in foals with a new cefquinome formulation: a field study. DTW., 116 (9): 316-20.
- Salmon, S. A.; Wattsm, J. L. and Yancey, R. J. (1996):** In vitro activity of ceftiofur and its primary metabolite desfuroylceftiofur, against organisms of veterinary importance. J. Vet. Diag. Inv., 8:322-336.
- Sheldon, I. M.; Bushnell, M.; Montgomery, J. and Rycroft, A. N. (2004):** Minimum inhibitory concentrations of some antimicrobial drugs against bacteria causing uterine infections in cattle. Vet. Rec., 25:155(13):383-387.
- Snedecor, G.W. (1969):** Statistical methods. 4th Ed. The Iowa state Univ. press, Ames., Iowa, USA., 91.
- Thomas, E.; Thomas, V. and Wilhelm, C. (2006):** Antibacterial activity of cefquinome against equine bacterial pathogens. Vet. Microbiol., 115(1-3):140-147.
- Tragesser, L. A.; Wittum, T. E.; Funk, J. A.; Winokur, P. L. and Rajala Schultz, P. J. (2006):** Association between ceftiofur use and isolation of *Escherichia coli* with reduced susceptibility to ceftriaxone from fecal samples of dairy cows. Amer. J. Vet. Res., 67:1696-1700.
- Venkanagouda; Krishnappa, G. and Upadhye, A.S. (1996):** Bacterial etiology of early chick mortality. Ind. Vet. J., 73: (3) 253-256.
- Walker, S. E.; Sander, J. E.; Cline, J. L. and Helton, J. S. (2002):** Characterization of *Pseudomonas aeruginosa* isolates associated with mortality in broiler chicks. Avian Dis., 46(4):1045-1050.

النشاط المضاد للبكتيريا لبعض مركبات السيفالوسبرون مع الإشارة الخاصة لتأثيرها على وزن ومناعة الكتاكيت

الفيومي المحصنة ضد مرض النيوكاسل

استهدفت هذه الدراسة معرفة حساسية ميكروب البروتيس ميرابيليس والقولون الإشيريكي والزائفة الزنجارية المعزولة من الحضانات وكتاكيت فيومي عمر يوم لثلاثة من المضادات الحيوية من مجموعة السيفالوسبرون وهي السيفرادين والسفتيفيور والسيفكينوم وتم حساب تركيزاتها المثبطة والقاتلة لهذه البكتيريا وتم أيضا دراسة تأثير هذه المضادات الحيوية على معدل نمو ونفوق الكتاكيت ودراسة تأثير هذه المضادات الحيوية على الجهاز المناعي للكتاكيت المحصنة بمصل النيوكاسل. وأتضح من الاختبار المعمل أن هذه الميكروبات كانت حساسة للسفتيفيور والسيفكينوم ولكن ميكروب القولون الإشيريكي والزائفة الزنجارية أكثر مقاومة لعقار السيفرادين. ووجد أن أقل معدل نمو كان في الكتاكيت التي لاتأخذ مضادات حيوية (105.7 ± 6.5 جم) تليها مجموعة الكتاكيت المحقونة بالسفتيفيور (162.5 ± 2.0 جم) وكان أعلى معدل نمو في المجموعة المحقونة بالسيفكينوم (183.5 ± 1.66 جم) عند عمر 30 يوم. ووجد أن أعلى نسبة نفوق كانت في المجموعة التي لاتأخذ مضادات حيوية تليها مجموعة الكتاكيت المحقونة بالسيفرادين تليها المجموعة المحقونة بالسفتيفيور ثم المجموعة المحقونة بالسيفكينوم. ومن هذه الدراسة تبين أن هذه المضادات الحيوية ليس لها تأثير مثبط للجهاز المناعي في الكتاكيت المحصنة ضد مرض النيوكاسل.